

In situ localization of matrix components

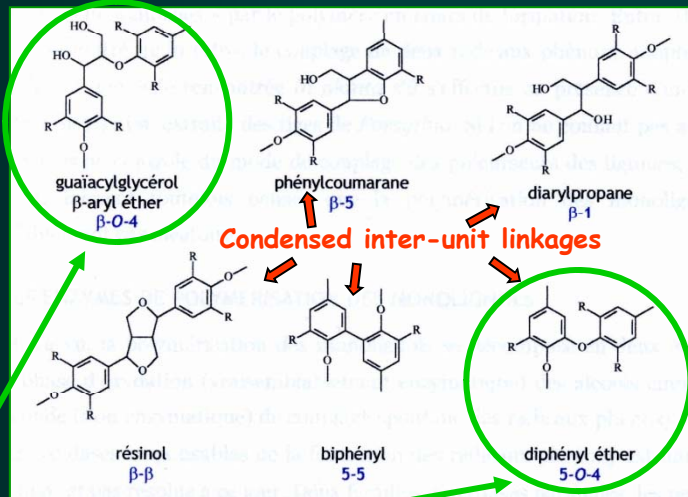
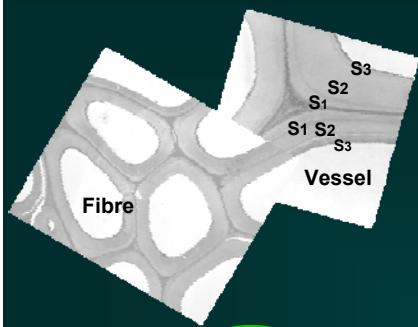
Lignins

- ◆ Ruel et al. **J Trace Microprobe Techniques** 1994, 12:247-265
- ◆ Joseleau and Ruel **Plant Physiol.** 1997, 114:1123-1133
- ◆ Joseleau, K. Ruel. . **CR. Acad. Sci.** Paris, 2004, 327: 809-816
- ◆ Joseleau et al. **Planta**, 2004, 219:338-345
- ◆ Joseleau and Ruel in **New Knowledge in Wood Quality**, K. Entwistle & J.C.F. Walker Eds., 2005, 103-113
- ◆ Joseleau and Ruel,, **Cellulose Chem Technol**, 2007, 41: 487-494



Wood cell wall components

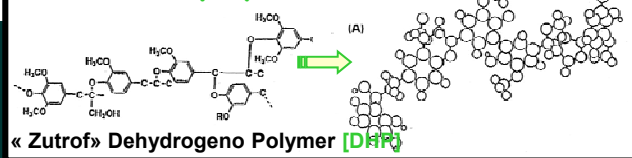
Lignin



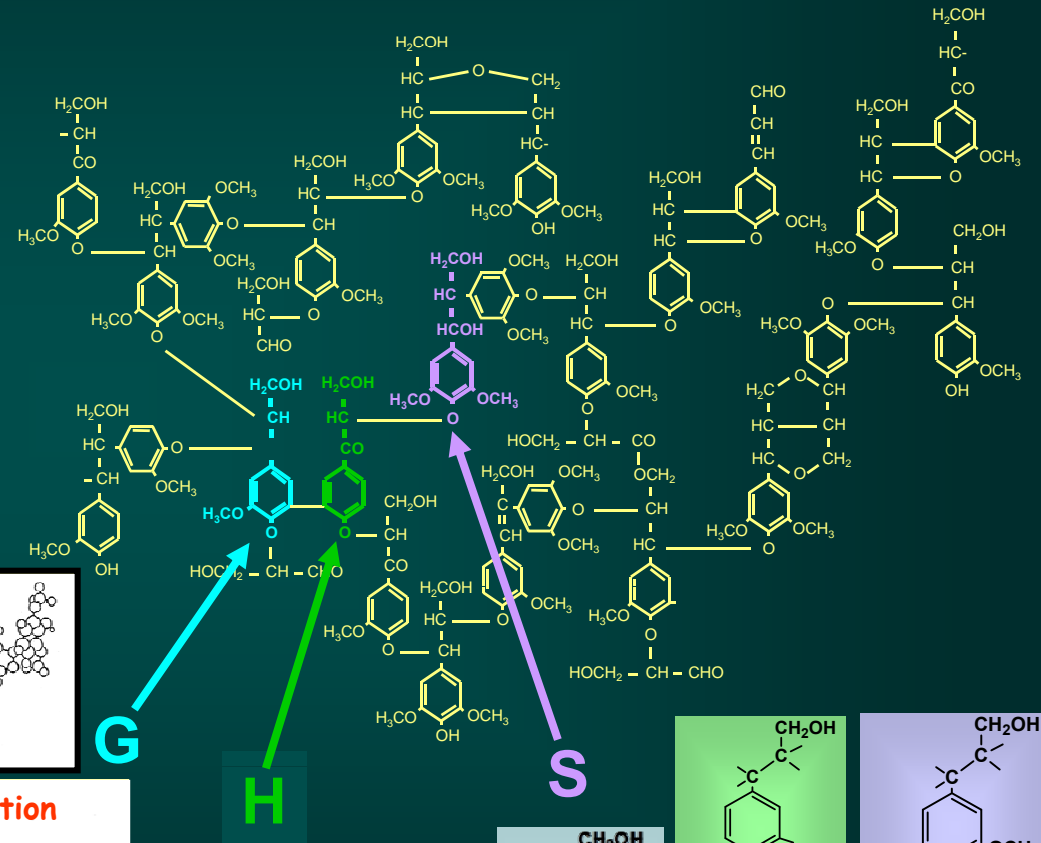
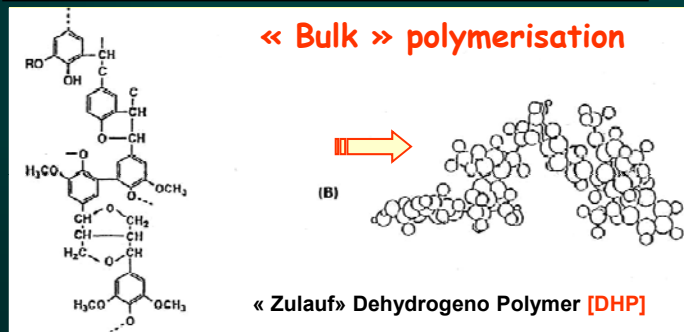
Non-condensed inter-unit linkages

Two modes of polymerization of the Monomers

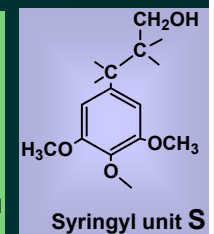
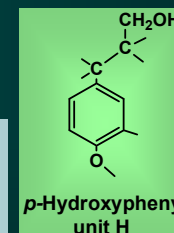
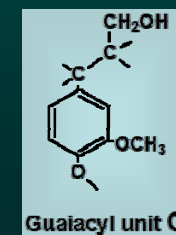
«endwise» polymerisation



« Bulk » polymerisation



Three basic Monomers



Antibodies from synthetic antigens (DHPs)

➤ 1) directed against monomer units

- A
- A
- A
- A

The specificity of our anti-lignin antibodies is directed against:

- Constitutive monomers (⇒ *composition*)
- Inter-unit linkages (⇒ *condensed / non-condensed*)
- Macromolecular conformation (⇒ *extended vs bulky*)

➤ 2) directed against

➤ 3) Specificity

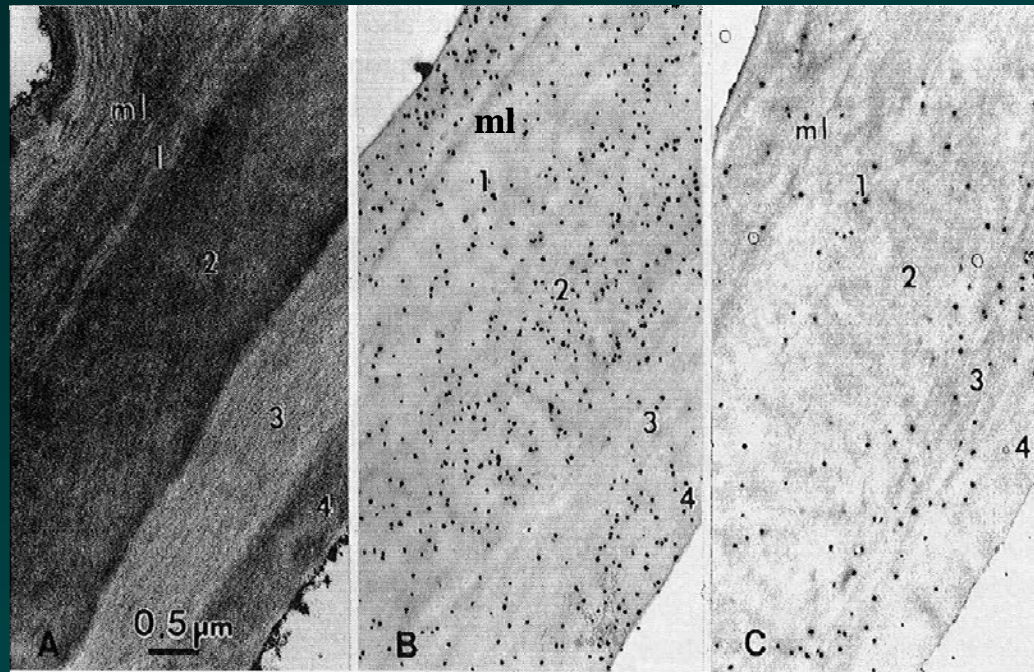
binding
TEM affinity tests*

Ruel et al. , J. Trace Microprobe, 1994
*Joseleau and Ruel, Plant Physiol., 1997
Ruel et al. CR Biol., 2004

TEM

Distribution of matrix components: Lignins

Vessels

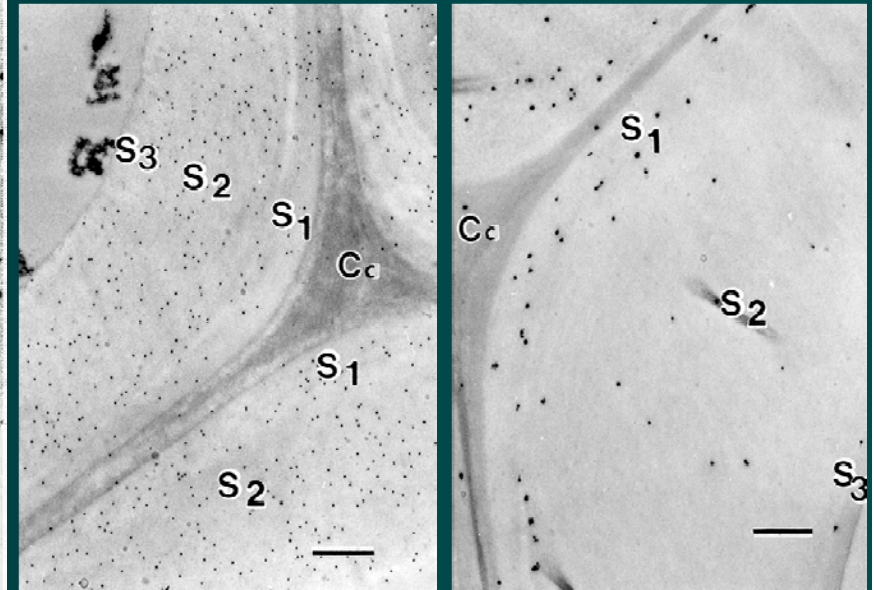


KMnO4

Non-condensed

Condensed

Fibers



Non-condensed

Condensed

☛ The distribution of the main lignin types (GS, G, S) is **different:**

☞ *From one tissue to another*

☞ *Inside the cell-wall itself: **topochemical microdomains***

☛ *Patterns are characteristic of cell type (Fiber/vessel)*

♦ Ruel et al. *Plant Biol.* 2002, 3: 2-8

♦ Joseleau and Ruel *Plant Physiol.* 1997, 114:1123-1133

♦ Ruel et al. *Maderas Cienc Tecnol*, 2006, 8:107-116

Katia Ruel



TEM

In Planta topochemical distribution of the main types of lignin in Poplar Wood

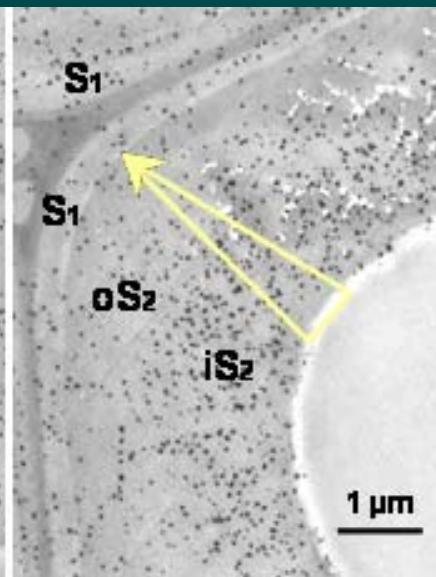
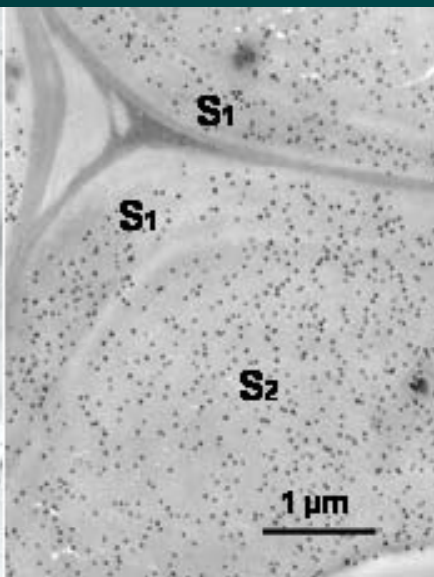
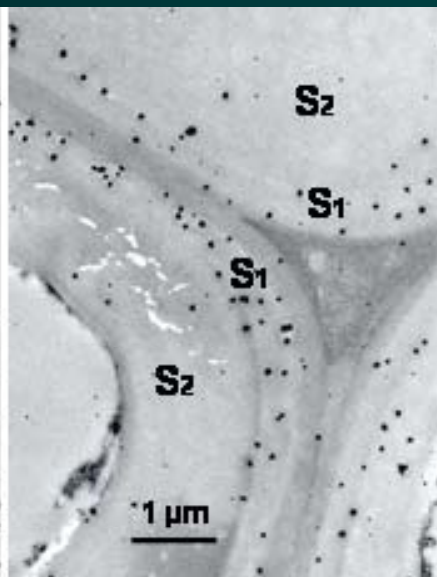
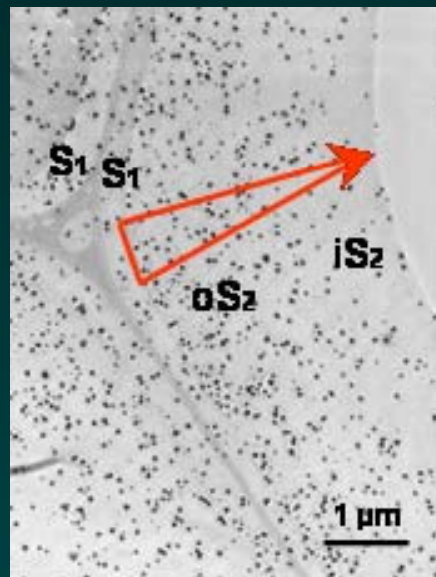
Condensed Homo-G

Condensed Mixed-GS

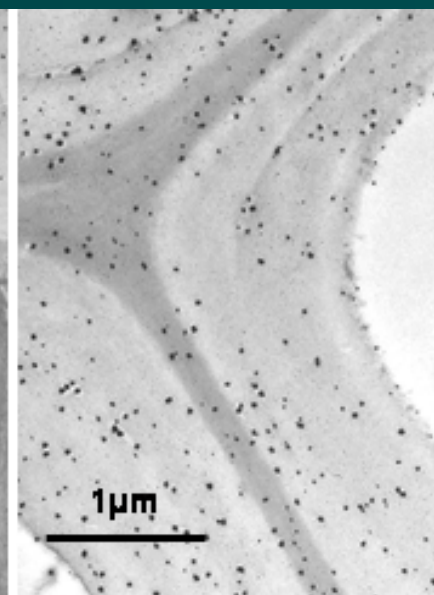
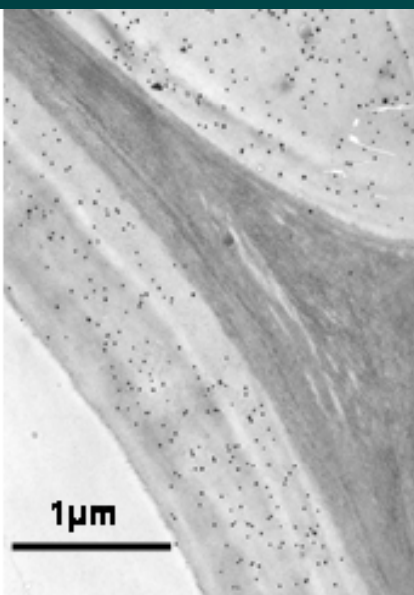
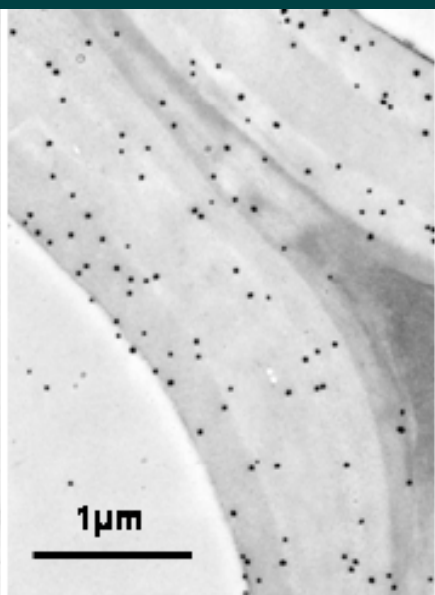
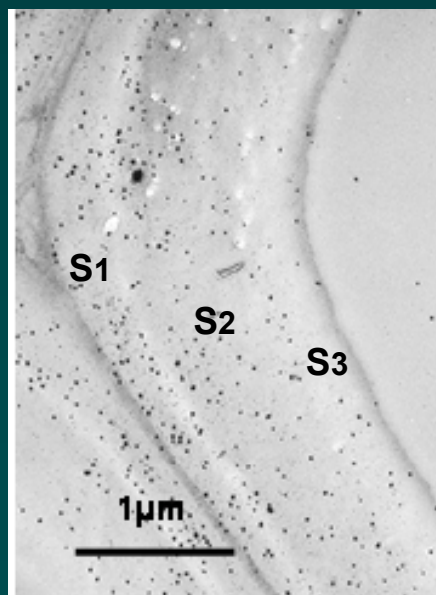
NonCondensed GS

NonCondensed S

FIBERS



VESSELS



Lignin topochemistry

- Immunolabelling demonstrates **regio-specificity** in Secondary Walls
- ***Condensed*** and ***Non-condensed*** Lignin epitopes are **differentially distributed** in Secondary Walls



Monolignol Polymerization is
Spatially and Temporally
controlled during cell Wall
Assembly

◆ Role of Matrix Components in Cell Wall Structure and Assembly ?

- Cells in development
- Transgenic plants
- Deconstructing agents

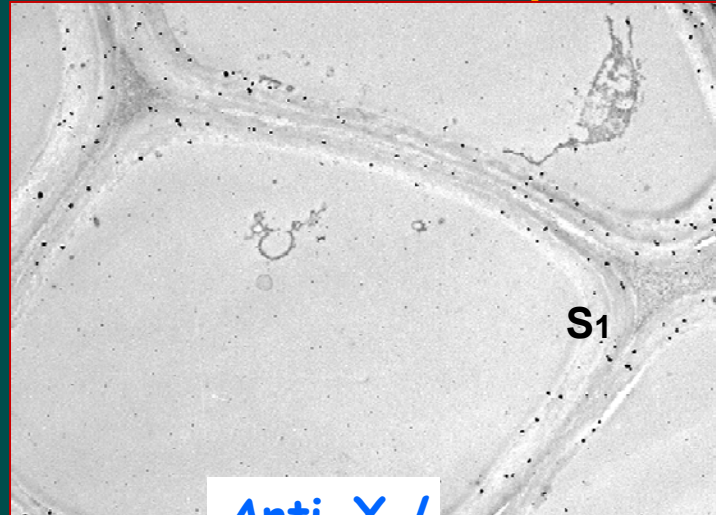
Role of matrix components in cell wall assembly

➡ Developing cells

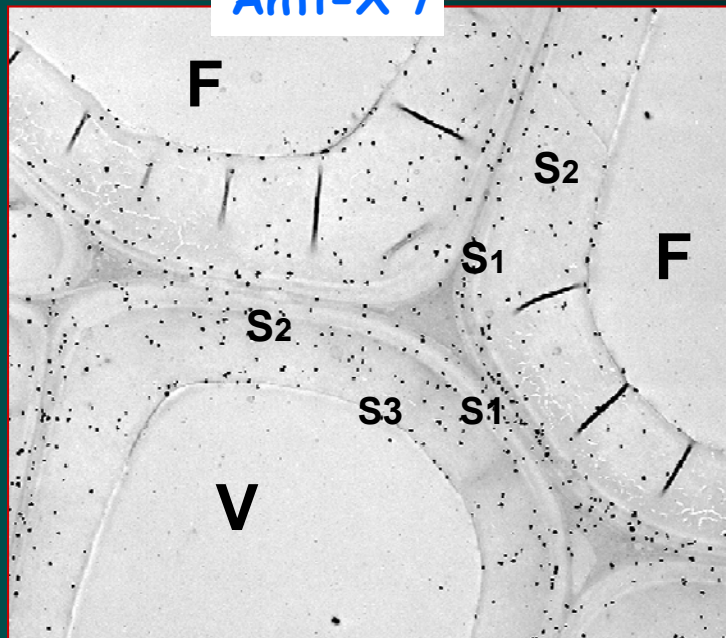
- ➡ Fate of xylans during cell wall development
- ➡ Fate of lignins during cell wall development

Fate of Xylans Deposition in Developing walls of Poplar wood

low-substitution Xylan



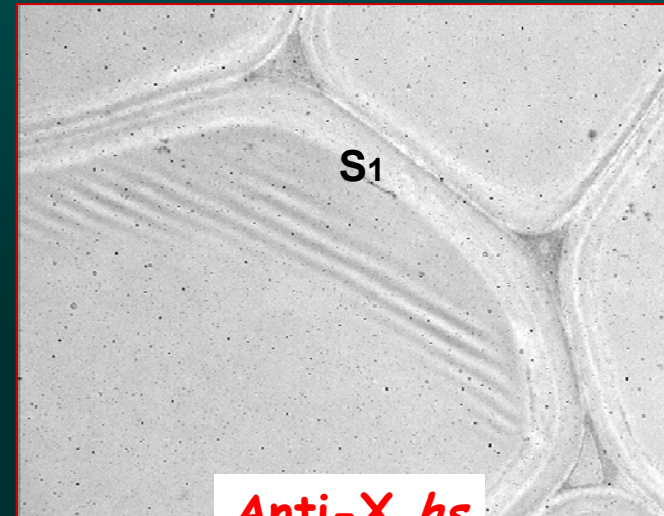
Anti-X /



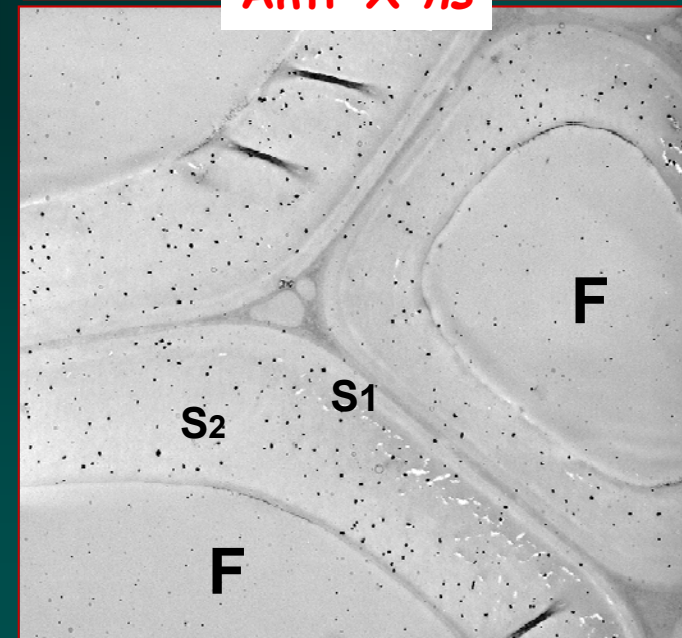
Young
stage

Mature
stage

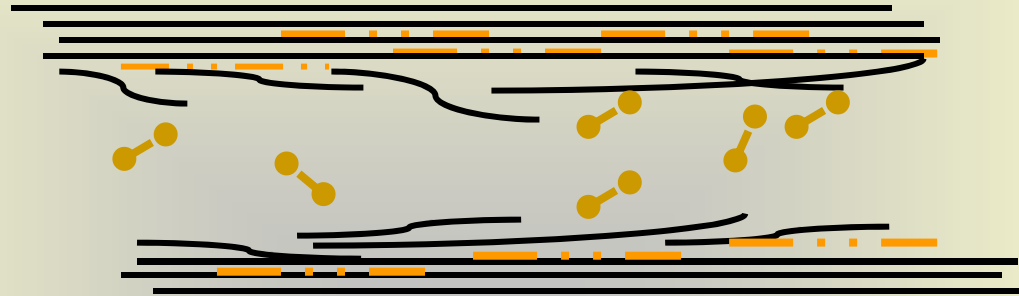
highly substituted xylan




Anti-X_{hs}




Sequential events in the deposition of **Xylans** in Hardwood



Event 1  Cellulose

 Linear Xylan (X l) *Event 2*

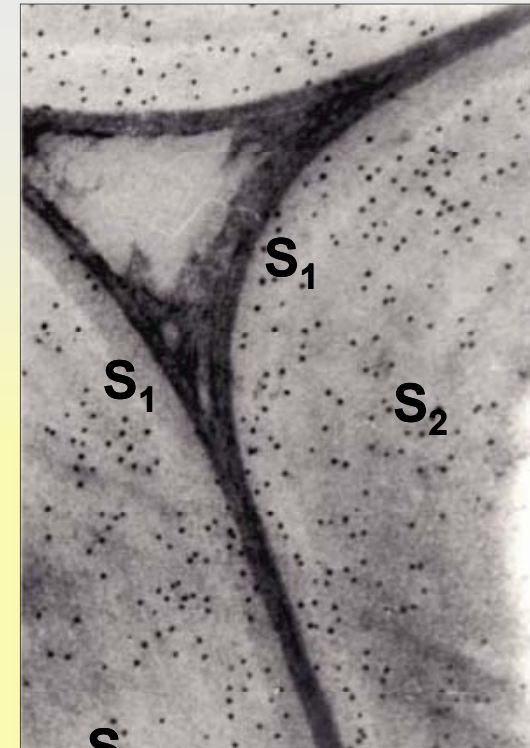
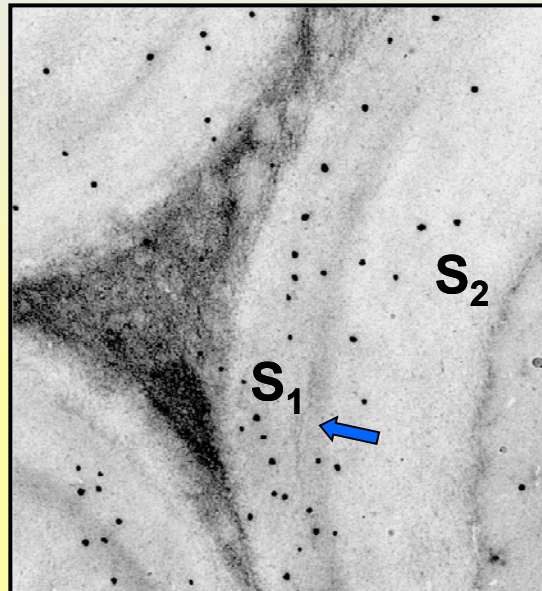
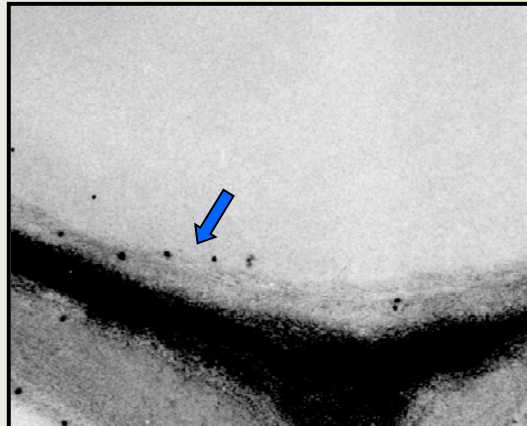
 Highly substituted
Xylan (X hs) *Event 3*

⇒ Recently, *Dammstrom---Salmen et al. (2009)* proposed that one fraction of xylans should be more strongly associated to cellulose

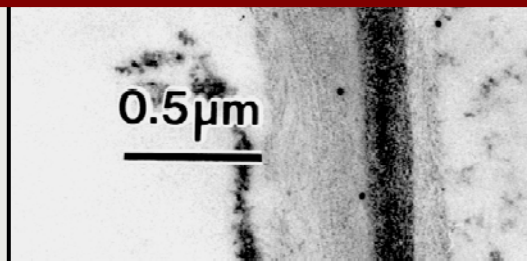
TEM

Fate of Lignins Deposition

Populus



• **Non-condensed** type of **lignin** is **early deposited** and takes part in the **cohesion** between lamellae



Immunolabelling of **non-condensed lignin sub-units**

Role of matrix components in cell wall assembly

➡ Genetically modified plants

➡ On the lignin biosynthesis pathway

The lignin biosynthetic pathway

Biosynthetic pathway of phenylpropanoids

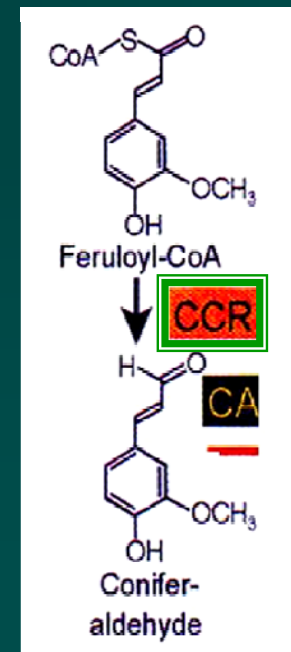
1/ Synthesis of phenolic aromatic nucleus

Shikimic acid \Rightarrow hydroxy cinnamic acids via phenylalanine

2/ \Rightarrow cinnamic alcohols
= *Monolignols*

3/ monolignols polymerisation


Lignin(s)



cinnamoyl-CoA
reductase
(CCR) down-regulation or
inhibition

Analytical data of lignins from CCR transformed plants

| Plant | Lignin content (Klason % weight) | Residual CCR activity (%) | %S | %G | S/G | Ferulic acid [μmol/(g KL) ⁻¹] | G-CHR-CHR2 |
|--|-------------------------------------|---------------------------|---------------|------|------|--|------------|
| Tobacco NN B-3-H | 22 | 100 | Non-condensed | | 0.9 | nd | nd |
| | 10.7 | 6 | Condensed | | 1.5 | nd | nd |
| | | | 21 | 79 | | | |
| <i>A.thaliana</i> WS As-CCR2 As-CCR7 Col0 SCCR1 irx4 | 17.0 | 100 | 28.2 | 71.6 | 0.39 | 1.7 | nd |
| | 10.0 | 19.2 | 34.1 | 65.5 | 0.52 | 5.0 | nd |
| | 13.0 | 24.4 | 36.3 | 63.4 | 0.57 | 6.1 | nd |
| | 18.3 | 100 | 28.1 | 71.2 | 0.40 | 1.7 | nd |
| | 11.7 | 0.0 | 27.6 | 71.3 | 0.39 | 18.5 | nd |
| | (-50%) | nd | nd | nd | nd | nd | nd |
| Poplar WT FS3 (Lepié et al. Plant Cell, 2007) | 20.7 | | | | 1.9 | [pmol/(mg Wood)] 75 | 2.2 |
| | 16.7 | | | | 1,5 | 111 | 3.4 |
| | | | | | | | |

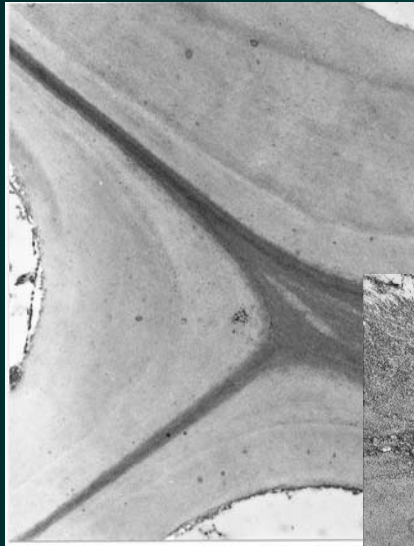
TEM

Consequences of CCR down-regulation in *Tobacco*

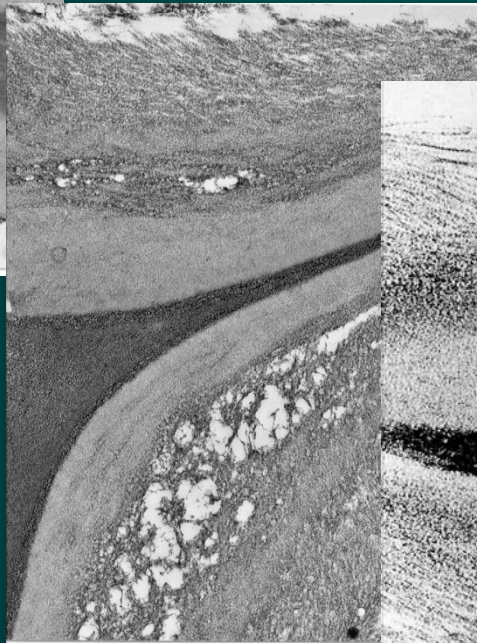
FIBRES

antisense

Nul mutant



WT



B3-H

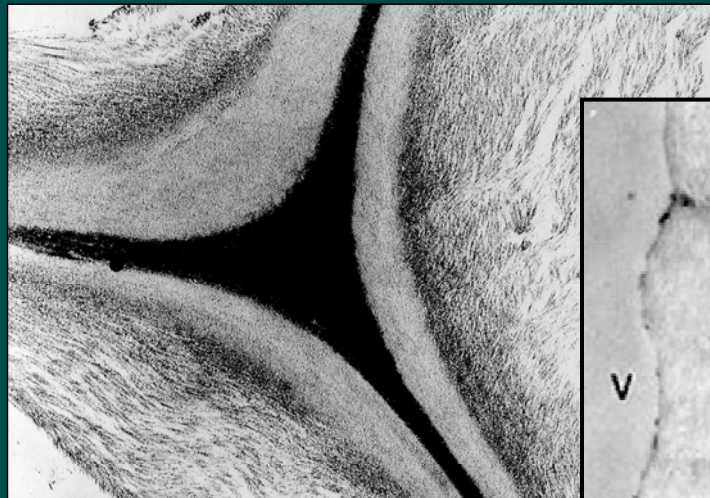


B3-1

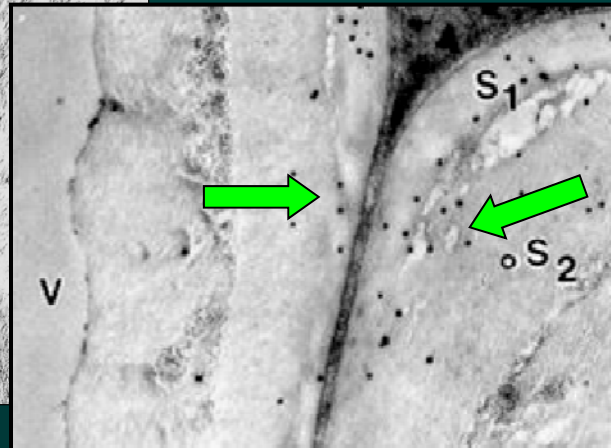
Dramatic
Loosening
of the
Secondary Wall (S₂)

TEM

Consequences of CCR down-regulation on lignin topochemical distribution

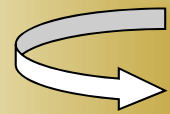
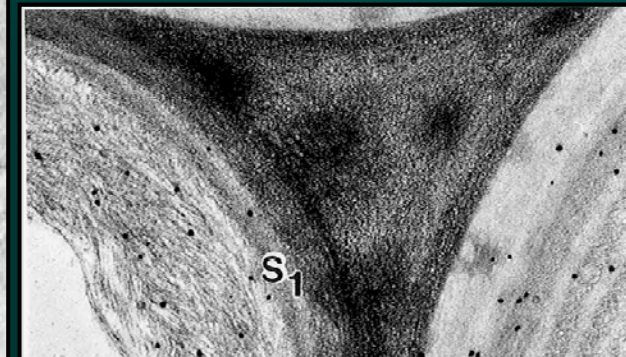


Non-condensed



Tobacco

Condensed



The non-condensed forms of lignin play a significant role in the organization of S2

Contributing to :

- ☛ The association of cellulose microfibrils
- ☛ The cohesion of the secondary cell wall.

TEM

Consequences of CCR down-regulation on lignin topochemical distribution

Leplé et al. Plant Cell 2007



Condensed G

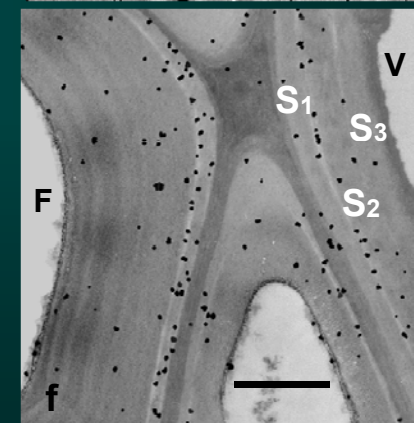
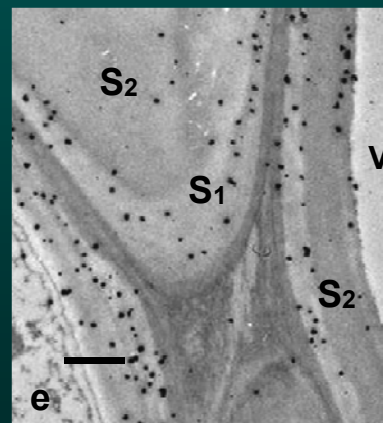
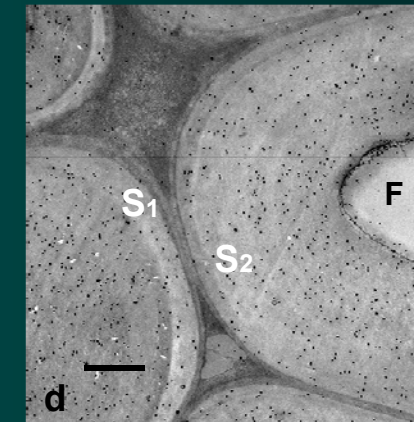
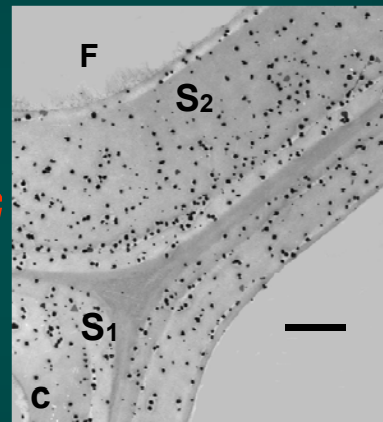
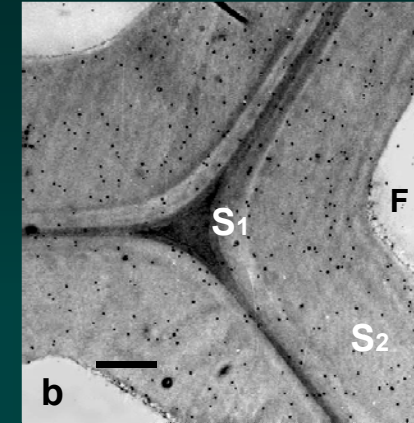
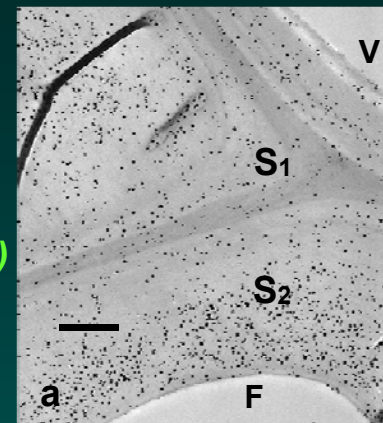
Non-Condensed
 α -O-8



Changes in the topochemical
distribution of lignin sub-units

Wild-Type

Transformed Poplar



Role of matrix components in cell wall assembly

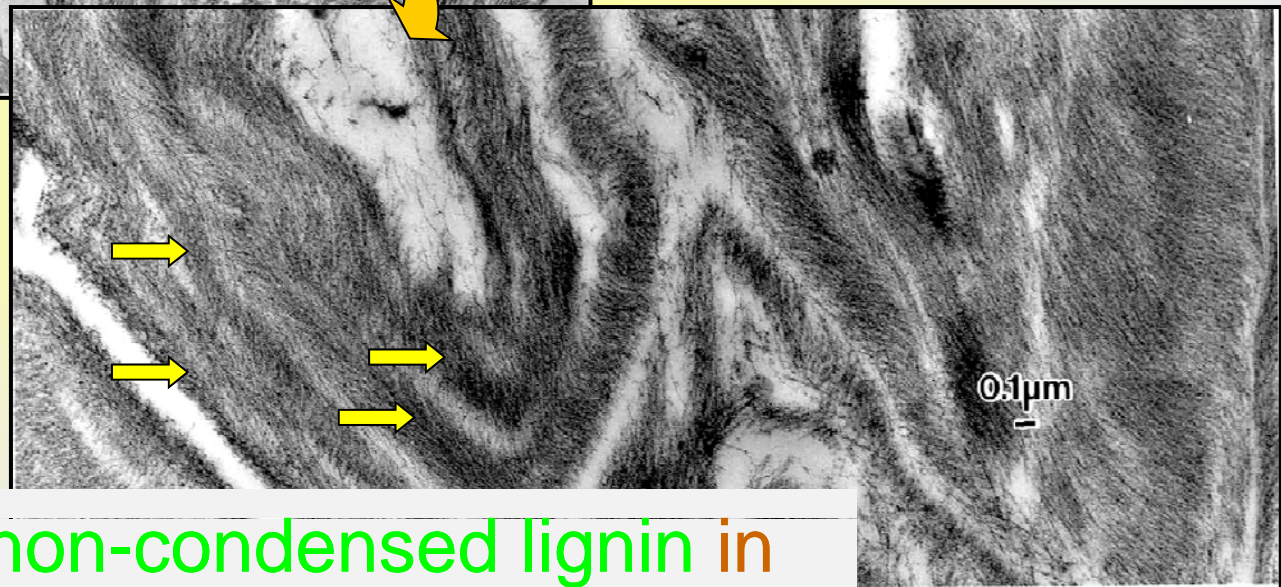
➡ Deconstructing agents

TEM

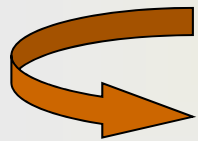


Unbleached Kraft pulp treated
with Manganese-Peroxidase
from *P.chrysosporium* I-1512

Sigoillot, JC, Ruel, K et al. (1997)
Holzforschung, 51:549-556



Role of non-condensed lignin in
cohesion between lamellae



**To Summarize the
Preceding
Observations.....**



Successive Steps in the Formation of Lignified Secondary Walls

➤ Biosynthetic Events

- i/ Newly synthesized CMFs [deposited in crystalline state]
- ii/ Low-substituted Xylans adsorbed on Cellulose
- iii/ Deposition of Lignin under *condensed* form
- iv/ Highly-sustituted Xylans deposit and serve as anchoring for *Non-condensed* Lignin
- vi/ *Non-condensed* Lignin covalently binds to uronic acid substituents of highly-substituted Xylans

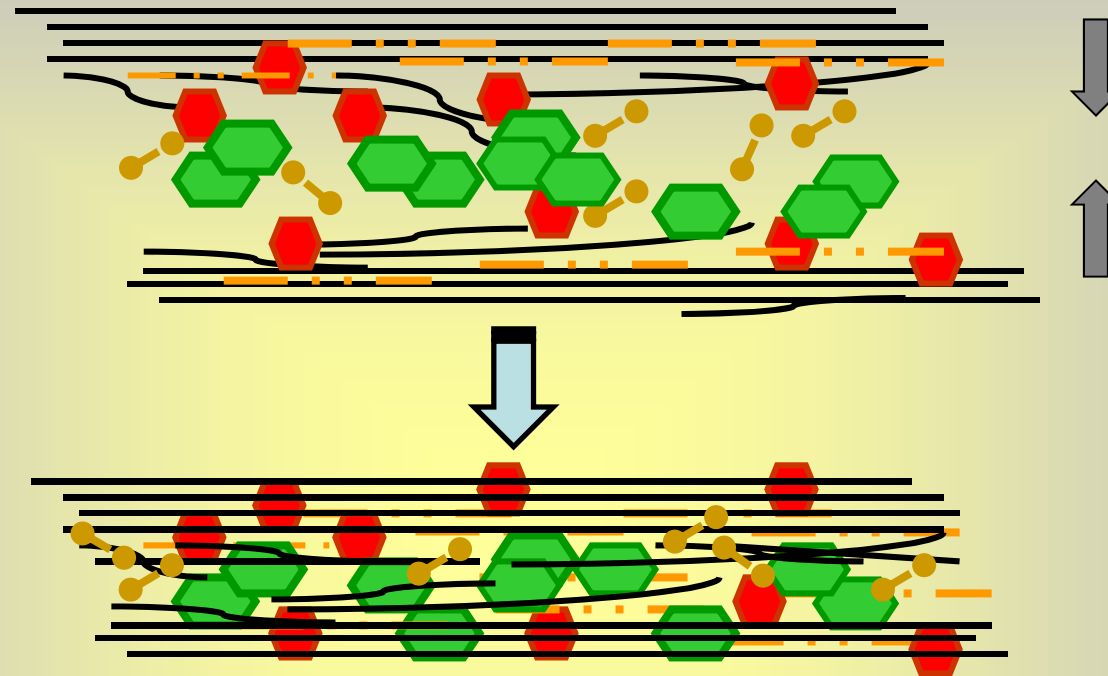
This mechanism repeats itself as S2 thickens.

➤ Consequences on Cell Wall Structure

The binding between *non-condensed* lignin and xylans induces tightening between CMF lamellae

➡ ➡ **Overall: *Cohesion + Compacity***




Dynamic Scheme of the Deposition of Cellulose, Hemicelluloses and Lignins during Secondary Wall Assembly





Plasmalemma

➤ **Cohesion**

➤ **Compacity**

Step 1  **Cellulose**
 Step 2  **Xylan *l***
 Step 3  **Condensed lignin**

Step 4  **Xylan *hs***
 Step 5  **Non-condensed lignin**

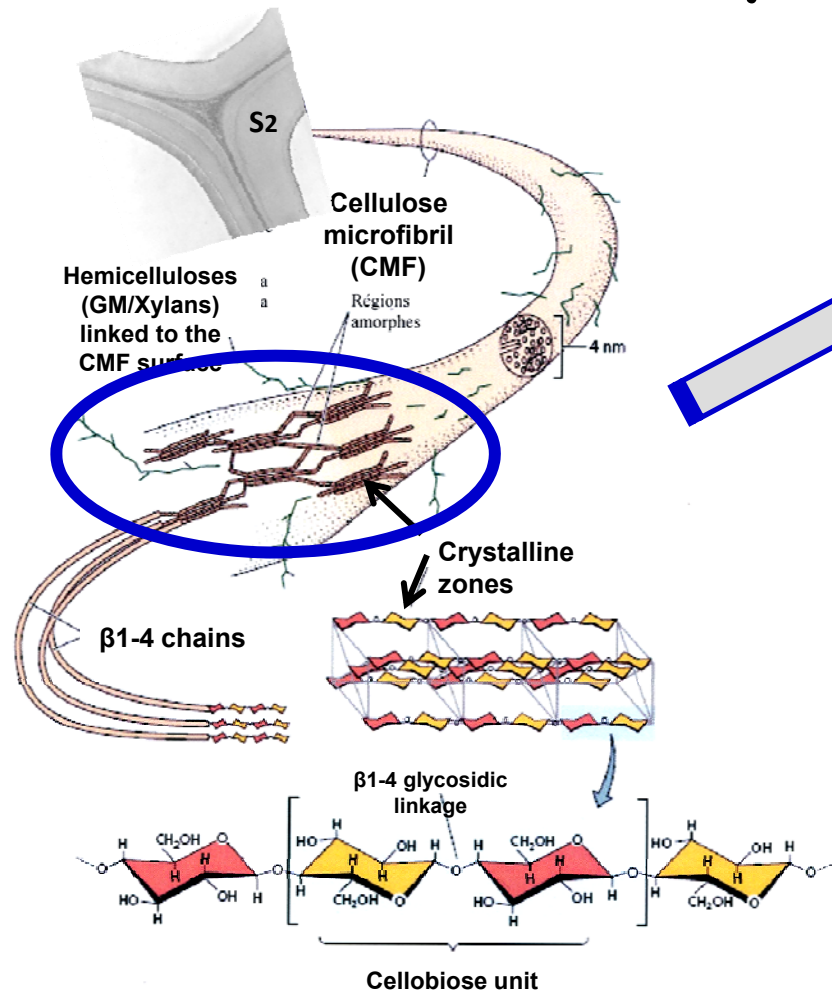
◆ The Cellulose Framework

➤ In planta Crystalline and Non-crystalline cellulose distribution

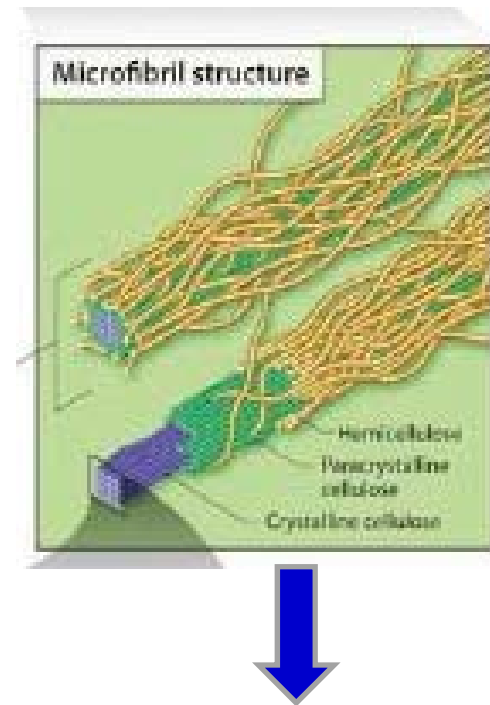
Cellulose microfibrils crystallinity in Woody Plants

- About half of the cellulose in fibres from Wood and crop plants is crystalline: the rest is disordered
- The proportion of crystalline cellulose in spruce wood has been estimated at 30%, equivalent to about 60% of the total cellulose. This fraction appears to be more than the 40% of cellulose I as defined conformationally by NMR (Fernandez..... jarvis 2011- PNAS)

Cellulose crystallinity in plant cell walls



From Chanliaud (2006)



Recent techniques confirmed that **well ordered cellulose forms the core** of each microfibril and much of the **less ordered cellulose is at the surface**, as has been suggested previously on the basis of NMR experiments (From Fernandes..... jarvis 2011-PNAS)

How crystalline and disordered parts fit together *in planta*....?

Visualization of Crystalline and Amorphous Cellulose in the Cell Walls

Cellulose-Binding Modules (CBMs)

Tools to probe Crystalline and Amorphous Cellulose *in situ*

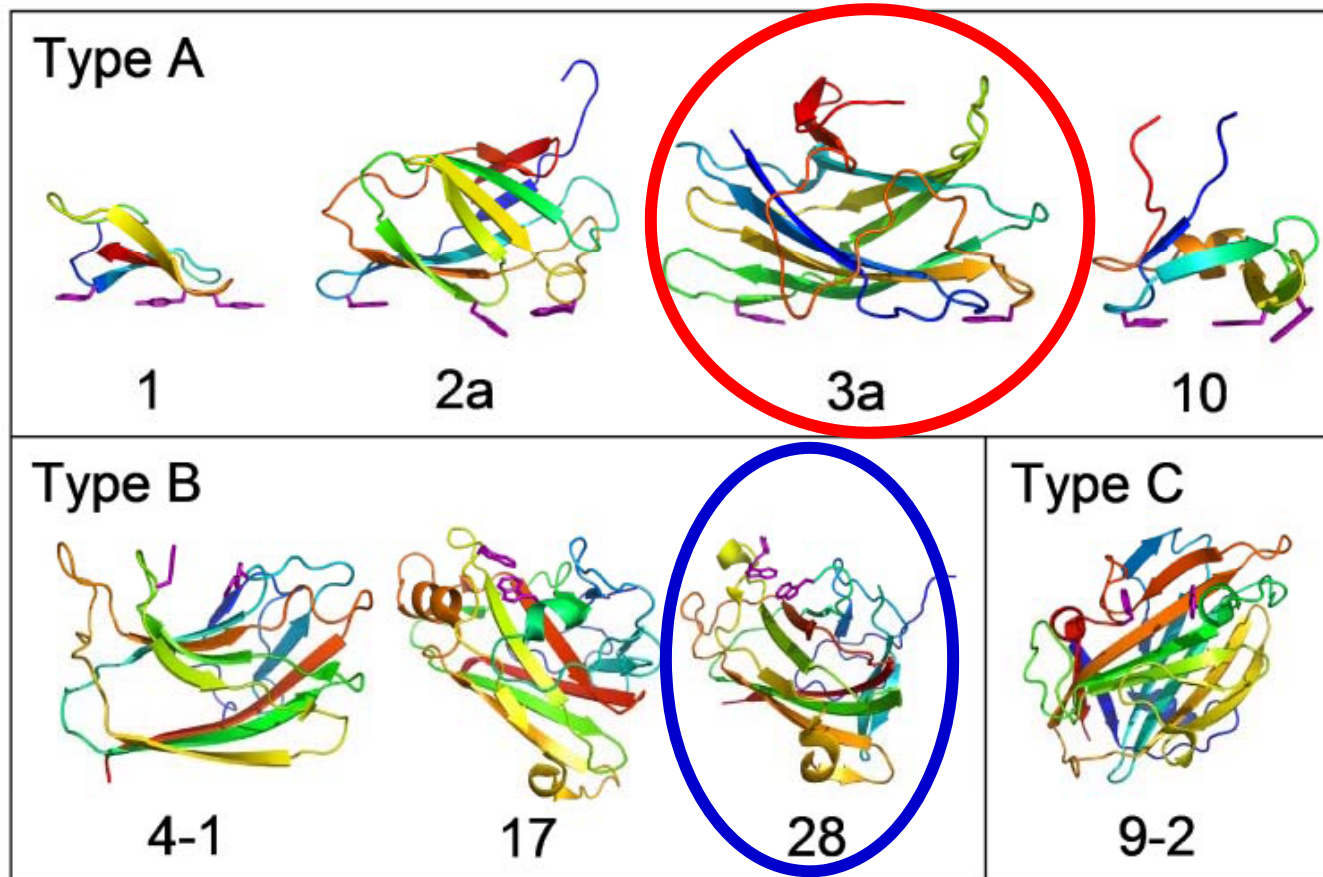
CBMs are a diverse set of **non-catalytic protein domains** that have a wide range of binding specificities towards cell wall polymers [Lehtio, 2003,].

Adapted as probes for light Microscopy

- Hildrén et al. (2003)

- Mc Cartney et al. (2004)

CBMs: Tools to probe Crystalline and Amorphous Cellulose *in situ*



Type A = surface binding CBMs which bind specifically to **crystalline cellulose**.

Type B = chain-binding CBMs specific for single chains of polysaccharides = **amorphous cellulose**

(Boraston et al. 2004)

Type C = end-binding CBMs specific for the ends of polysaccharides or oligosaccharides.

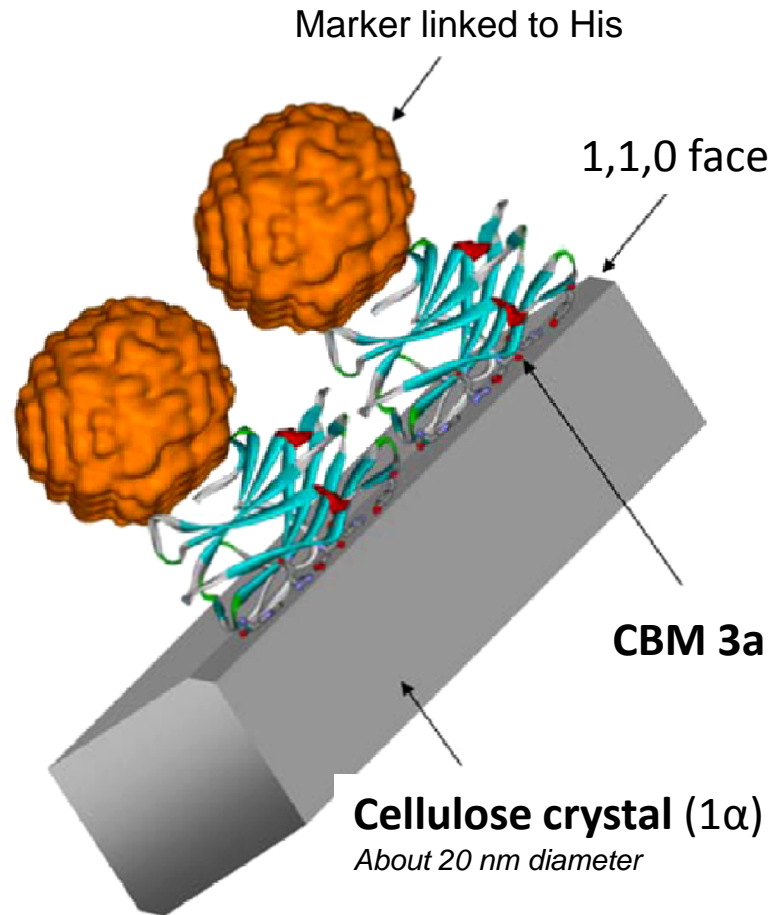


Fig. 1 A schematic model of cellulose/CBM/QD bio-assembly. The recombinant CBM protein specifically binds to the 1,1,0 surface of the cellulose 1 α crystal. The QDs are anchored on CBM proteins by the his-tags fused at both *N*- and *C*-termini. QDs, CBMs, and cellulose are depicted in scale based on their published X-ray structures. The cellulose crystal model is simplified (showing fewer glucan chains) to fit into the picture. The actual size of *Valonia* cellulose crystals used in this study is about 20 nm in diameter

From Xu et al. (2009) *Cellulose* 16:19-26

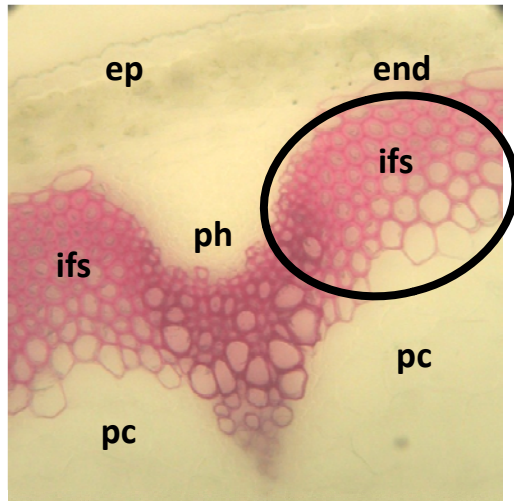
CBMs binding

Case of *Valonia* cellulose crystalline microfibrils, it has been in-situ demonstrated a **systematic binding to the two hydrophobic planar (110) faces of the cellulose microfibrils**

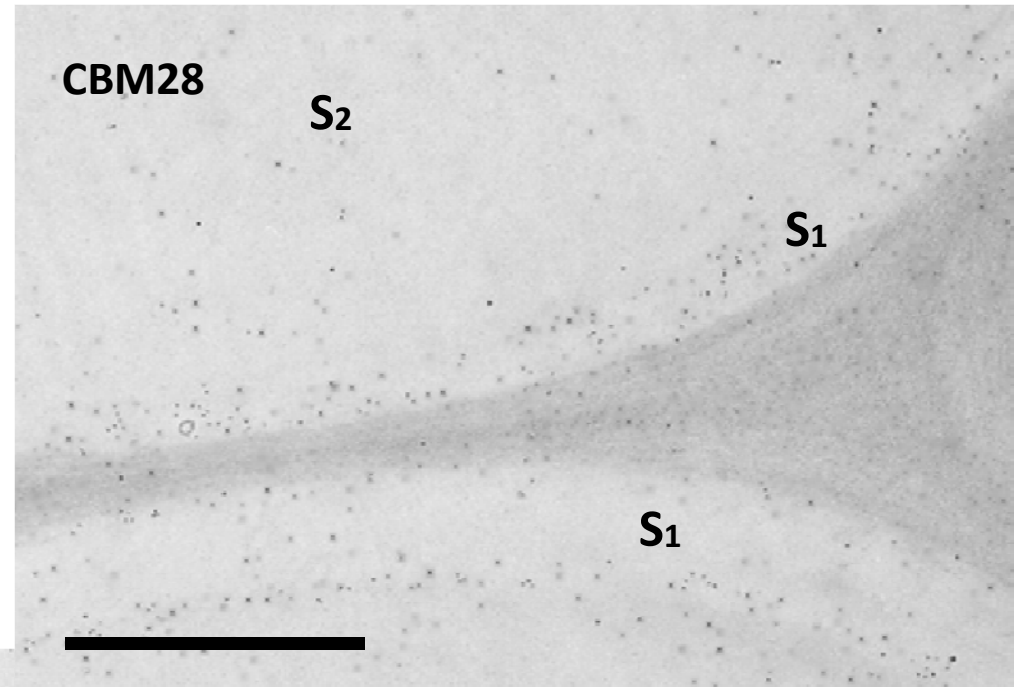
has been fluorescently labeled and **used to label crystals as well as plant tissue** (Ding et al., 2006; Porter et al., 2007; Liu et al., 2009; Xu et al., 2009).

????Dagel et al. « In situ imaging of single carbohydrate-binding modules on cellulose microfibrils ». *J. Phys. Chem. B* 2011, 115, 641

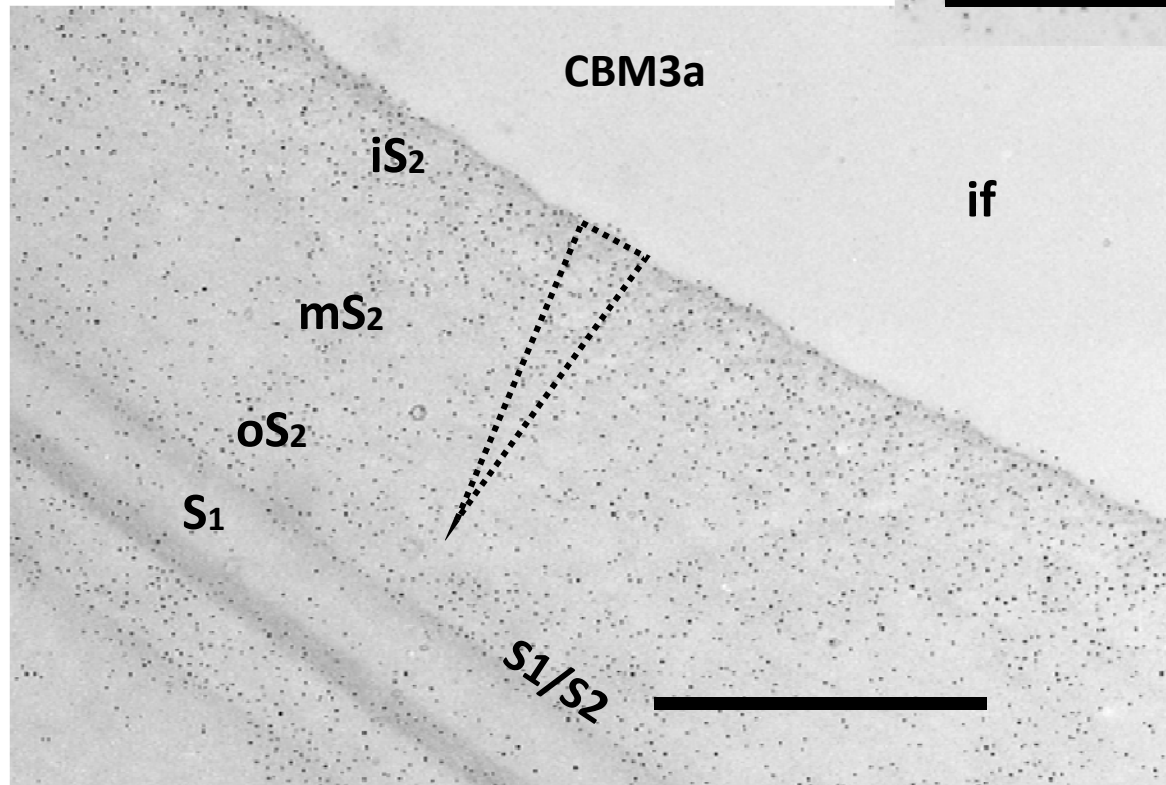




Crystalline cellulose



Amorphous cellulose



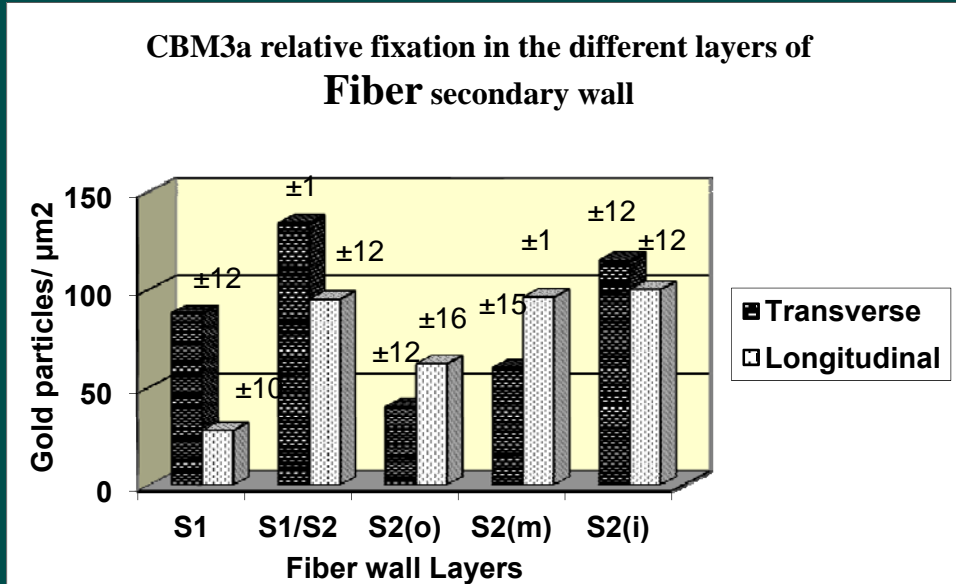
Distribution of crystalline and amorphous cellulose domains in lignified fibers (*A. thaliana*)

[Ruel, Nishiyama, Joseleau. *Plant Sci.* 2012
"Crystalline and Amorphous Cellulose in the
Secondary Walls of *Arabidopsis*"]

Katia Ruel

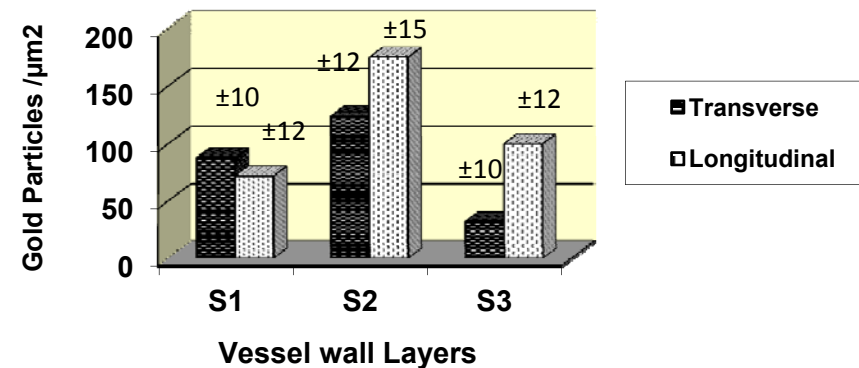


Quantitative Evaluation of CBM3a fixation across transverse and longitudinal sections



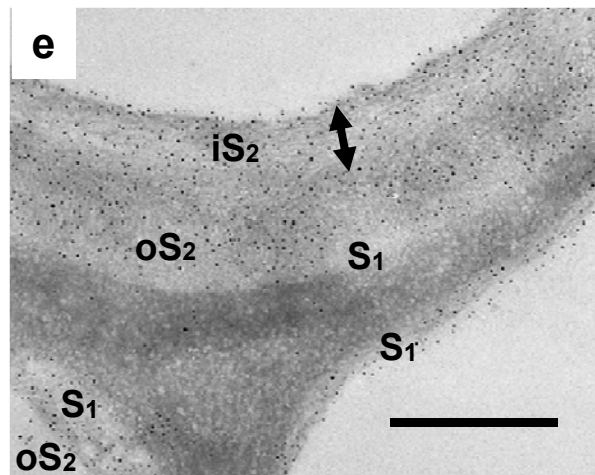
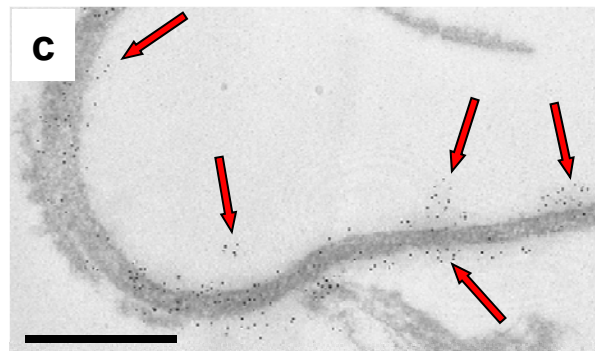
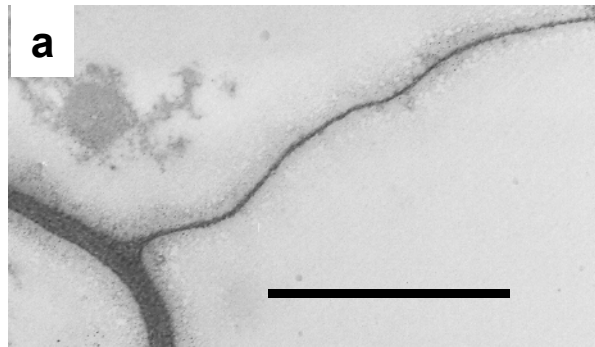
[Ruel, Nishiyama, Joseleau. *Plant Sci.* 2012
"Crystalline and Amorphous Cellulose in the
Secondary Walls of Arabidopsis"]

CBM3a relative fixation in the different layers of **Vessel** secondary wall

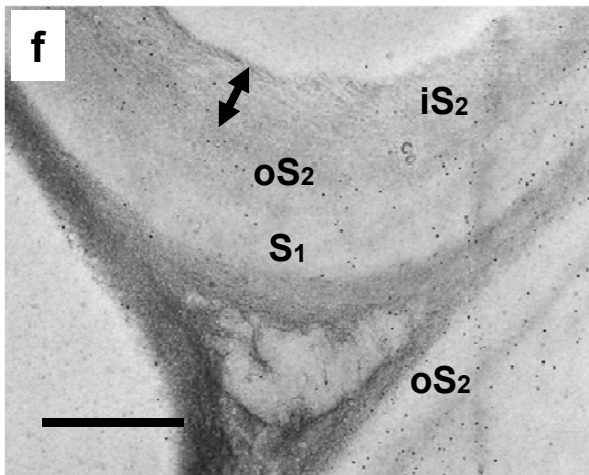
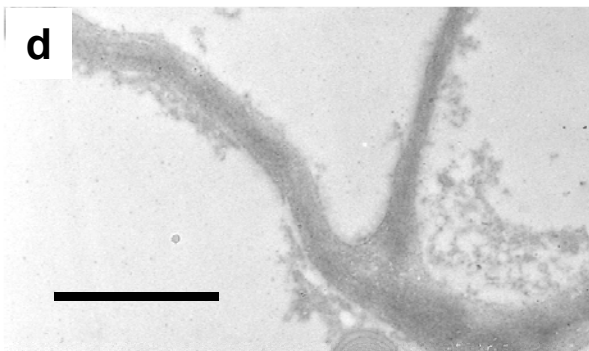
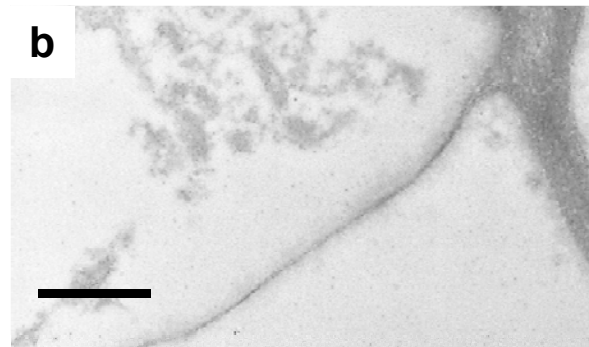


Mapping of Cellulose Deposition during Cell Wall Development

Crystalline Cellulose (CBM3a)



Amorphous Cellulose (CBM28)



Cellulose
first
deposits
under
Crystalline
Structure

[Ruel, Nishiyama, Joseleau. *Plant Sci.* 2012 "Crystalline and Amorphous Cellulose in the Secondary Walls of Arabidopsis"]

70



Conclusion

- 1/ Cellulose first deposits under *Crystalline* (or *High Crystallinity*) state
- 2/ *Amorphous* Cellulose then appears progressively, due to:
 - * **irregularities** in the biosynthesis process
 - * **growth stress** constraints [*internal stress*]
 - * **post-synthesis destructuration** (Endogenous enzymes; mechanical action) [*external stress*]

**Viewed at the Nano-scale , the Wood Cell
Walls are Complex Hierarchical
Structures**

**They display Multiple Organizations defining
the Diversity and Variability of Cell Types in
Adaptation to Specific Physiological
and Mechanical Functions**



Special Thanks to Prof. Em. Jean-Paul JOSELEAU

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For transformed Tobacco
 For CCR mutants of *A. thaliana*
 For CAD down-regulated poplar
 For down-regulated Tobacco
 For transformed Poplar
 For pulp samples
 For lignin thioacidolysis
 For computerized studies on lamellae
 For the gift of CBMs

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